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**Introduction**

In the evolving health management paradigm,1-4 the general regulation of the immune system as well as the enhancement of specific immune functions have become a growing point of interest, and rightly so. Many health problems result from the inability of the immune system to stop a disease process in its initial stage. This paper will review the scientific evidence for the immunomodulatory effects of blue-green algae and some of the demonstrated effects of blue-green algae on health and disease.

The human body is constantly being exposed to foreign organisms such as bacteria, viruses, fungi, and parasites, all of which coexist to a certain degree in the skin, the mouth, the respiratory tract, the intestinal tract, and the genital tract. Some microorganisms are essential for optimal health, and the healthy human body is well-equipped to keep such organisms from becoming a problem. However, when the natural barriers are compromised, or when we are exposed to more highly infectious organisms, serious disease may result. This includes not only acute infectious diseases, but also chronic inflammatory and autoimmune diseases. Optimal support of the immune system is important for prevention of and intervention with diseases with microbiological involvement, whether acute illness or chronic degenerative disease. Inflammation sets the stage for chronic disease, and for the initiation and progression of cancer. Enormous research efforts are currently pursuing nutritional and botanical intervention of inflammatory processes.

**Blue-green Algae as Food**

Blue-green algae (cyanobacteria) are among the most primitive life forms on Earth. Their cellular structure is a simple prokaryote. They share features with plants, as they have the ability to perform photosynthesis. They share features with primitive bacteria because they lack a plant cell wall. Interestingly, they also share characteristics of the animal kingdom as they contain on their cellular membrane complex sugars similar to glycogen. Among blue-green algae we find both edible and toxic species, adapted to almost any of the most extreme habitats on Earth, including deep-sea vents, hot springs, and Antarctica’s ice. Edible blue-green algae, including *Nostoc*, *Spirulina*, and *Aphanizomenon* species have been used for food for thousands of years. Habitats with sufficient algae growth include the Pacific Ocean near Japan and Hawaii, and large freshwater lakes, including Lake Chad in Africa, Klamath Lake in North America, Lake Texcoco in Mexico, and Lake Titikaka in South America. African and American natives recognized the value of including blue-green algae in their diet and stored dried algae for year-round use and trade.

Still today, edible blue-green algae are a nutrient-dense food. As for any other crop, differences exist with regard to harvest procedures, quality control for contaminating species, adherence to proper processing to preserve nutrients from degradation, and storage conditions. The nutrient content depends on the location and environment in which the algae was grown as altitude, temperature, and sun exposure can greatly affect lipid and pigment composition. *Spirulina* is an algae species grown at sea or in man-made ponds, and the mineral profile will differ from fresh-water algae such as *Aphanizomenon*. Algae grown in a natural environment will differ from algae grown in canals or tanks due to differences in aeration, nutrient circulation and availability, and degree of competition with other algal species. As we learn more about the phytoceutical components of different blue-green algae species, the optimal growth conditions for obtaining optimal yields can be determined.

The nutrient profile is subject to much variation between habitats and harvest procedures which influences the content of vitamins and antioxidants delivered in the final product. Certain features are common to all blue-green algae, including a high content of bioavailable amino acids and minerals, including zinc, selenium, and magnesium. Industrial standards still vary greatly in terms of documenting product composition to the consumer. However, blue-green algae have the appeal of being a raw, unprocessed food, rich in carotenoids, chlorophyll, phycocyanin, and many other bioactive components.

**Beyond Nutrition**

Among blue-green algae, many species have documented biomodulatory effects. This paper will review scientific evidence for immunomodulatory effects of blue-green algae and some of its demonstrated effects on health and disease. The research studies span the use of the whole algae of various species in both human and animal studies, as well as in vitro studies on algae extracts and purified compounds (Table 1).

**Table 1. Research On Blue-Green Algae As Biomodulators**

|  |  |  |
| --- | --- | --- |
| **Studies on** | **Route of administration** | **Compounds investigated** |
| Human | Oral consumption\* | Whole algae |
| Chicken | Oral consumption | Whole algae |
| Rodents | Oral consumption\*\* | Whole algae |
| Injection | Isolated fractions |
| Injection | Purified compounds |
| In Vitro | Added to media | Isolated fractions |
| Added to media | Purified compounds |
| *\*) Humans: oral dose was 1.5 – 2.8 grams per day for adult subjects* |
| *\*\*) Mice: oral dose varied up to 800 mg/kg* |

**EFFECTS OF BLUE-GREEN ALGAE ON INNATE (NON-SPECIFIC) IMMUNITY**

Several studies have examined the use of whole blue-green algae in the context of the normal functioning immune response. In our lab, one study using oral doses of 1.5 grams of the blue-green algae *Aphanizomenon flos-aquae* on healthy human volunteers revealed it slightly decreases the phagocytic activity of polymorph nucleated cells in vitro.5 This may indicate an anti-inflammatory, rather than anti-phagocytic effect on human neutrophils.

In a study looking at the phagocytic function of cat bronchoalveolar macrophages in vitro, the percentage of cells that phagocytosed cells increased when they were exposed to a water-soluble extract of *Spirulina* for two hours.6 The number of particles ingested by the phagocytic macrophages did not change when compared to control cultures.

In another study, mice were fed a *Spirulina*-supplemented diet (10% of the dry weight of food) for ten weeks, and the ability of peritoneal macrophages to ingest latex particles was evaluated in vitro. The results of this study showed a slight increase in the percentage of phagocytic cells (4.6%; from 91.3 to 95.9%).7 A similar effect was observed in chickens.8

In addition, murine peritoneal macrophages exposed in vitro to a hot-water extract of *Spirulina* for 24 hours secreted a substance, (speculated to be IL-1), which induced thymocyte proliferation.7 In the same study, the ability of spleen cells extracted from algae-fed mice to proliferate in response to mitogens was examined in vitro. These experiments showed that splenic cells isolated from algae-fed mice proliferated more when exposed to certain mitogens compared to control mice.

The effect of blue-green algae on non-specific immunity has also been examined at the level of natural killer (NK) cell activity. Using a standard chromium release assay, splenic leukocytes from chickens fed blue-green algae were shown to exhibit greater anti-tumor cell activity when compared to those of control animals.8 The authors speculate that blue-green algae may increase NK cell activity via the production of cytokines such as interferon.

In a study designed to investigate the mechanism behind the immunostimulatory effect of blue-green algae on the human monocyte/macrophage cell line THP-1, a crude extract of the blue-green algae *Aphanizomenon flos-aquae* was used to stimulate the cell line. The extract was half as potent as LPS in activating NF-kB, and the purified molecule is ten times more potent than LPS (Pasco, manuscript in press). The molecule responsible for this activation has been identified as a novel polysaccharide.9

Thus, multiple studies on whole blue-green algae in humans, mice, rats, cats, and chickens have demonstrated an effect on phagocytosis, NK cell function, and inflammation. Some differences exist in the data, including the mild reduction of phagocytic activity in humans after algae consumption, in contrast to the increase of phagocytosis among bronchoalveolar macrophages. The cell types and experimental set-ups vary, and further studies are needed to establish the exact biochemical mechanisms involved.

**EFFECTS OF BLUE-GREEN ALGAE ON SPECIFIC IMMUNITY**

Hayashi et al 7 examined the effect of an algae-supplemented diet on the ability to build a specific immune response to sheep red blood cells. After immunizing mice (either once to measure the primary response or twice for the secondary response), they found that mice fed with the algae-supplemented diet showed increased numbers of splenic IgM anti-body-producing cells when compared to control animals. Interestingly, this finding only held true for the primary immune response, as the IgG antibody production in the secondary immune response was hardly affected. In experiments involving chickens, there were no differences observed in anti-sheep red blood cell antibodies during primary responses, while antibody titers for the secondary response in algae-fed chickens were augmented compared to control animals.8 The differences may reflect the anatomical differences between the rodent and chicken immune systems.

Hayashi et al 10 examined other antibody classes such as IgA and IgE in the context of mice orally immunized with a crude shrimp extract. They found that whereby both IgA (intestinal) and IgE (in serum) levels increased with antigen challenge, only IgA levels showed a greater enhancement in secretion with concurrent treatment with *Spirulina* extract (five-week feeding regimen).10 From this study they concluded that blue-green algae does not seem to induce or enhance food allergic IgE-dependent reactions. Furthermore, they suggest that when ingested along with or before a potential antigenic threat, blue-green algae may enhance IgA antibody levels to protect against food allergies.

Along the same lines, further studies have suggested that blue-green algae may inhibit mast cell-mediated type I allergic reactions and even the anaphylactic reaction in rats.11,12 By injecting a blue-green algae extract intraperitoneally (100-1000mg/g body weight) one hour prior to an allergic challenge, mortality induced by the anaphylactic compound 48/80 was decreased, local allergic reaction activated by anti-dinitrophenyl (anti-DNP) IgE was inhibited, and serum histamine levels were decreased. In vitro experiments from this group provided similar results.

The effects of blue-green algae on IgE-production and allergic reactions are encouraging, and warrant further studies in humans.

**Table 2. Immuno-Modulatory and Anti-Inflammatory Effects of Whole Blue-Green Algae**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Algae Species** | **Introduced as:** | **Test Species** | **Effects** | **Reference** |
| *Spirulina* sp. | Food | Human | Reversal of tobacco-induced oral cancer | Mathew et al, 1995 |
| Food | Mouse | Proportional reduction of IgE, increase of IgA | Hayashi et al, 1998 |
| Food | Mouse | Increased phagocytic activityIncreased spleen cell proliferationIncreased antibody production | Hayashi et al, 1994 |
| Food | Chicken | Increased phagocytic activityIncreased NK cell-mediated anti-tumor activityIncreased antibody production | Qureshi et al, 1996 |
| Extract | In vitro, cat | Increased phagocytic activity | Qureshi & Ali, 1996 |
| IP injection | Rat | Inhibition of mast cellsDecrease in local allergic reactionDecrease in serum histamine levelsReduced allergy-induced mortality | Kim et al, 1998Yang et al, 1997 |
| *Aphanizomenon flos-aquae* | Food | Human | Increased transient recruitment of NK cells into tissueIncreased mobilization of T and B cells into bloodMild modulation of PMN-mediated phagocytic response | Jensen et al, 2000 |
| Food | Rat | Decreased serum levels of arachidonic acid | Kushak et al, 2000 |
| Food | Rat | Source of linolenic acid (omega-3)**Increased serum levels of EPA and DHA** | Kushak et al, 2000 |
| Extract | In vitro, rat | Activation of macrophages (NF-kappaB, cytokines) | Pasco, in press |

**EFFECTS OF BLUE-GREEN ALGAE ON LEUKOCYTE TRAFFICKING**

Much attention with regards to dietary modulation of the immune system has been given to stimulating activity of various immune cell types such as the phagocytic activity of macrophages, or the tumoricidal activity of natural killer cells. However, immune cell trafficking and the recruitment of immune cells from the systemic circulation are of equal importance. A recent study by Jensen et al 5 involving humans demonstrated that the blue-green alga *Aphanizomenon flos-aquae* was able to trigger within two hours the migration of nearly 40% of the circulating natural killer cells. This effect was significantly more pronounced in long-term consumers than in naÏve subjects. In the same study, *Aphanizomenon flos-aquae* was also shown to stimulate the mobilization of T and B lymphocytes. This effect appeared cell-type specific since no changes were observed on polymorph nucleated cells.

**ANTI-INFLAMMATORY PROPERTIES OF BLUE-GREEN ALGAE**

Blue-green algae in general contain a significant amount of carotenoids, namely beta carotene, lycopene, and lutein, providing it with good antioxidant properties. By their quenching action on reactive oxygen species, antioxidants carry intrinsic anti-inflammatory properties. However, blue-green algae also contains specific anti-inflammatory properties as a result of their high phycocyanin content. Phycocyanin is a photoharvesting pigment that provides the intense blue color in blue-green algae. It can constitute up to 15% of the dry weight of a blue-green algae harvest. C-phycocyanin is a free radical scavenger,26 and has significant hepatoprotective effects.27 Phycocyanin was shown to inhibit inflammation in mouse ears 28 and prevent acetic acid induced colitis in rats.29 The anti-inflammatory effect seemed to be a result of phycocyanin to inhibit the formation of leukotriene B4, an inflammatory metabolite of arachidonic acid.28

In a study performed in rats, the blue-green algae *Aphanizomenon flos-aquae* was also shown to decrease the plasma level of arachidonic acid.30 *Aphanizomenon flos-aquae* contains significant amounts of the omega-3 alpha-linolenic acid. Omega-3 fatty acids have been shown to inhibit the formation of inflammatory prostaglandins and arachidonate metabolites. Since *Spirulina* contains significant amounts of omega-6 gamma-linolenic acid, the anti-inflammatory properties of *Spirulina* must be due to different biochemical pathways.

**Table 3. Bio-modulatory Effects of Purified Compounds from Blue-Green Algae**

|  |  |  |  |
| --- | --- | --- | --- |
| **Species** | **Compound** | **Effects** | **References** |
| *All blue-green algae* | C-Phycocyanin | Anti-inflammatory (reduces leukotriene B4) | Romay 1999 |
| Free radical scavenger | Bhat & Madyastha 2000 |
| Selective inhibition of COX-2 | Reddy et al, 2000 |
| Reduced tissue damage in acetic acid-induced colitis | Gonzalez et al, 1999 |
| Hepatoprotective effect | Vadiraja et al, 1998 |
| *Spirulina* | Calcium Spirulan (Ca-sp) | Selectively inhibits penetration of virus into host cell (Herpex Simplex, human cytomegalovirus, measles, mumps, Influenza A, HIV-1) | Hayashi et al, 1996 |
| Reduces lung metastasis of melanoma cells by inhibition of tumor cell invasion of basal membrane | Mishima et al, 1998 |
| Cyanovirin-N | Irreversible inactivation of several strains of HIV (inhibited cell-to-cell and virus-to-cell fusion) | Boyd, 1997 |
| Extracellular products | Promotion of lactic acid bacteria growth in vitro | Parada et al, 1998 |
| *Aphanizomenon flos aquae* | Unknown | Induces apoptosis in some human tumor cell lines | Jensen, msp in prep |
| Polysaccharide | Stimulate the macrophage activity | Pasco et al, in press. |
| *Lyngbya lagerheimii Phormidium tenue* | Sulfolipid | Inhibits syncytium formation upon HIV infection | Gustafson et al, 1989 |
| *Phormidium tenue* | Digalactosyl diacylglycerols | Inhibition of chemically induced skin tumors | Tokuda et al, 1996 |

**ANTI-VIRAL EFFECTS**

As part of its program aimed at discovering new anti-tumor and anti-viral agents in natural sources, the National Cancer Institute isolated extracts of blue-green algae (Lyngbya lagerheimii and Phormidium tenue) that were found to protect human lymphoblastoid T cells from the cytopathic effect of HIV infection. Upon further investigation, a new class of HIV inhibitory compounds called the sulfonic acid-containing glycolipids were isolated; the pure compounds were found to be strikingly active against the HIV virus in the p24 viral protein and syncytium formation assays.13 Since this discovery, there has been further investigation into other species of blue-green algae for compounds with anti-viral properties. Some compounds worthy of mention include a protein called cyanovirin-N which appears to irreversibly inactivate diverse strains of the HIV virus and to inhibit cell-to-cell and virus-to-cell fusion.14 Other studies using a water-soluble extract of blue-green algae have found a novel sulfated polysaccharide, calcium spirulan (Ca-SP), to be an antiviral agent. This compound appears to selectively inhibit the penetration of enveloped viruses (Herpes simplex, human cytomegalovirus, measles virus, mumps virus, influenza A virus, and HIV-1) into host cells, thereby preventing replication. 15-17 A review of anti-HIV activity of extracts from blue-green algae has been recently published.18

**ANTI-CANCER EFFECTS**

An early study on blue-green algae’s cancer-preventive properties in humans was performed on tobacco-induced oral leukoplakia.19 Mathew et al found that oral supplementation with Spirulina fusiformis resulted in complete regression of 57% of subjects with homogenous leukoplakia. After discontinuation of Spirulina supplementation, almost half of the complete responders developed recurrent lesions.

In other studies, extracts of blue-green algae have been used to treat cancer in animal models. In one model, ingestion of an extract of Spirulina and Dunaliella was shown to inhibit chemically-induced carcinogenesis in hamster buccal pouches.20,21 Earlier studies often attributed the anti-cancer effect of algae to its content in carotenoids since beta-carotene has been shown to have an effect similar to that of algae extract. Amore recent study, however, showed that the sulfated polysaccharide mentioned above, Ca-SP, appears to inhibit tumor invasion and metastasis.22 Both the in vitro and in vivo effects of Ca-SP suggest that the intra-venous administration of Ca-SP reduces the lung metastasis of melanoma cells by inhibiting the tumor invasion of the basement membrane. Awater-based extract of Aphanizomenon flos aquae containing high concentrations of phycocyanin inhibited the in vitro growth of one out of four tumor cell lines tested, indicating that at least some tumor cell types may be directly sensitive to killing by phycocyanin (Jensen et al, manuscript in preparation). Another fresh-water blue-green algae, Phormidium tenue, contains several diacyl-glycerol compounds which effectively inhibited chemically-induced skin tumors in mice.23 In addition, Spirulina was shown to have a modulatory effect on hepatic carcinogen metabolizing enzymes.24

Of major interest to ongoing research in inflammation as well as breast cancer is the finding that C-phycocyanin selectively inhibits COX-2, but has no effect on COX-1. 25 The COX enzymes are involved in prostaglandin synthesis. Since COX-2 is over-expressed in many breast cancer cells, and inhibition of COX-2 leads to a markedly reduced tumor growth and blocks angiogenesis, the finding that phycocyanin specifically interferes with this pathway holds promise.

**BLUE-GREEN ALGAE AS A BIOMODULATOR**

Besides their effects on the immune system, blue-green algae have also been reported to modulate other systems and improve metabolism. In the past few years increasing attention has been given to the study of the therapeutic effects of blue-green algae. The anecdotal claims for such effects are numerous. Although there is limited data from controlled animal or clinical studies, such claims include improvement in condition of Alzheimer’s patients, overall enhancement of immune response, improvement in fibromyalgia, control of hypertension, alleviation of depression and chronic fatigue, increased stamina, healing of internal and external lesions, increased mental acuity, and general improvement in overall well-being. This last section will review the scientific evidence supporting the therapeutic effects of blue-green algae.

**EFFECTS ON METABOLISM**

Several reports from different labs have shown that certain species of blue-green algae have cholesterol-lowering effects in animal and human models. In feeding experiments in rats, two studies have reported that the elevation in total cholesterol, LDL, and VLDL cholesterol in serum caused by cholesterol feeding was reduced when the high cholesterol diet was supplemented with 16% and 5% blue-green algae, respectively.31,32 In addition, Kato found that adipohepatosis induced by a high fat and high cholesterol diet was cured rapidly when the diet was supplemented with algae.31 Investigations into the mechanism of this phenomenon led to the finding that the algae-fed group showed a statistically significant increase in the activity of lipoprotein lipase, a key enzyme in the metabolism of triglyceride-rich lipoproteins.33 The hypocholesterolemic effect of blue-green algae was also observed in humans in a study conducted on 30 patients with mild hyperlipidemia and mild hypertension.34 Patients took 4.2 grams of algae or placebo per day, and were observed for two months. At the end of the study, patients taking the algae showed a significant reduction of LDL-cholesterol (p<0.05) compared to the control group. LDL cholesterol increased back to baseline levels after administration of the algae was discontinued. In addition to lowering LDL cholesterol levels, the atherogenic index (a measure of fat deposition in arteries) declined significantly after four weeks of algae consumption.

In a recent study by Kushak et al, rats were fed the blue-green alga Aphanizomenon flos-aquae and total cholesterol level was monitored. After 43 days, cholesterol levels were significantly decreased when compared to the control group.30 Although Aphanizomenon flos-aquae contains a significant amount of the omega-3 polyunsaturated linolenic acid, the effect on cholesterol levels seemed unrelated to the lipid content of the diets. Kushak et al 30 proposed that the hypocholesterolemic effect of Aphanizomenon flos-aquae may be due to its chlorophyll content which was shown to stimulate liver function and decrease blood cholesterol.35

In a double-blind crossover study involving human patients, supplementing the diets of obese outpatients with 2.8 grams of blue-green algae three times daily over a four week period resulted in a statistically significant reduction of body weight.36 In a study measuring the effect of blue-green algae on glucose levels in diabetic rats, the water-soluble fraction was found to be effective in lowering the serum glucose level at fasting, while the water insoluble fraction suppressed glucose levels at glucose loading.37 In another study investigating the effect of the blue-green alga Aphanizomenon flos-aquae on rat intestinal mucosal digestive enzymes, it was observed that this alga specifically inhibited the activity of maltase and sucrase in a dose-dependent manner.38 Furthermore, this decrease in enzymatic activity was accompanied by a dose-dependent decrease in blood glucose.

The overall conclusion is that blue-green algae may have benefits on lipid and sugar metabolism, as well as liver function. Further human studies are needed to address the feasibility of using blue-green algae in conjunction with cholesterol-lowering medication.

**Table 4. Biomodulatory Effects of Whole Blue-Green Algae on Metabolism**

|  |  |  |  |  |
| --- | --- | --- | --- | --- |
| **Algae Species** | **Introduced as** | **Test Species** | **Effects** | **Reference** |
| *Aphanizomenon* | Food | Rat | Reduction of cholesterol | Kushak et al, 2000 |
| Food | Rat | Reduction of blood glucose levels | Drapeau et al, 2001 |
| Food | Human | Modulation of brain activity (EEG) | Walker & Valencia, 1999 |
| *Spirulina* | Food | Human | Reduced body weight | Becker et al, 1986 |
| Food | Rat | Reduction of cholesterol | Kato et al, 1984 |
| Food | Rat | Increased activity of lipase | Iwata et al, 1990 |
| Food | Rat | Reduced glucose levels | Takai et al, 1991 |
| Food | Rat | Inhibition of maltase and sucrase | Kushak et al, 1999 |
| Food | Mouse | Modulation of carcinogen metabolic enzymes | Mittal et al, 1999 |
| Food | Mouse | Modulation of lead toxicity | Shastri et al, 1999 |
| Food | Rat | Increased iron status during pregnancy and lactation | Kapoor & Mehta, 1998 |
| *Nostoc* | Food | Rat | Reduction of cholesterol | Hori et al, 1994 |

**OTHER EFFECTS OF BLUE-GREEN ALGAE**

Other research studies on blue-green algae consumption deserve mention. Many reports exist in the literature on its antimicrobial effects. The secretion of anti-microbial substances is an important part of the competition for ecological niches in the natural environment. However, an interesting caveat exists. In one study, Spirulina was cultured in vitro, and the extracellular medium was shown to stimulate the growth of lactic acid bacteria.39 If the growth-promoting substance(s) exist in sufficient amounts intracellularly, blue-green algae may play a role in vivo by supporting friendly gut bacteria. This leads to other facets of health including gut health and nutrient absorption. On that note, consumption of Spirulina was shown to support the iron status and hemoglobin of rats during pregnancy and lactation.40 Spirulina fusiformis had a significant protective effect against lead-induced toxicity in rats.41 Finally, a report by Valencia et al has presented evidence that Aphanizomenon flos-aquae accelerates recovery from mild traumatic brain injury.42

**CONCLUSION AND SUMMARY**

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**REFERENCES**

1. Eisenberg D, Davis R, et al. Trends in alternative medicine use in the United States. 1990-1997. *JAMA*. 1998:280(18):1569-1575.
2. Astin J. Why patients use alternative medicine. *JAMA*. 1998;279(19):1548-1553.
3. Sobel D. Rethinking medicine: improving health outcomes with cost-effective psychosocial interventions. *Psychosomatic Medicine*. 1995;57:234-244.
4. Furnham A. Forey J. The attitudes, behaviors, and beliefs of patients of conventional vs complementary (alternative) medicine. *J Clini Psych*. 1994;50:458-469.
5. Jensen GS, Ginsberg DI, et al. Consumption of *Aphanizomenon flos aquae* has rapid effects on the circulation and function of immune cells in humans. JANA. 2000;2(3):50-58.
6. Qureshi M, Ali R. *Spirulina platensis* exposure enhances macrophage phagocytic function in cats. *Immunopharmacol Immunotoxicol*. 1996;18(3):457-463.
7. Hayashi O, Katoh T, et al. Enhancement of antibody production in mice by dietary *Spirulina platensis*. *J Nutr Sci Vitaminol*. 1994;40:431-441.
8. Qureshi M, Garlich J, et al. Dietary *Spirulina platensis* enhances humoral and cell-mediated immune function in chickens. *Immunopharmacol Immunotoxicol*. 1996;18(3):465-476.
9. Pugh N, Ross SA, ElSohly HN, ElLohly MA, Pasco DS. Isolation of three high molecular weight polysaccharides with potent immunostimulatory activity from *Spirulina platensis*,*Aphanizomenon flos-aquae* and *Chlorella pyrenoidosa*. *Planta Medica*. In Press.
10. Hayashi O, Hirahashi T, et al. Class-specific influence of dietary *Spirulina platensis* on antibody production in mice. *J Nutr Sci Vitaminol*. 1998;44(6):841-851.
11. Kim H, Lee E, et al. Inhibitory effect of mast cell-mediated immediate-type allergic reactions in rats by *Spirulina*. *Biochem Pharmacol*. 1998;55(7):1071-1076.
12. Yang H, Lee E, et al. *Spirulina platensis* inhibits anaphylactic reaction. *Life Sci*. 1997;61(13):1237-1244.
13. Gustafson K, Cardellina II J, et al. AIDS-antiviral sulfolipids from cyanobacteria (blue-green algae). *J National Cancer Institute*.1989;81:1254-1258.
14. Boyd M. Protein isolated from blue-green algae inactivates HIV. *Antimicrob Agents Chemother*. 1997;41:1521-1530.
15. Hayashi K, Hayashi T, et al. A natural sulfated polysaccharide, calcium spirulan, isolated from *Spirulina platensis*: in vitro and ex vivo evaluation of anti-herpes simplex virus and anti-human immunodeficiency virus activities. *AIDS Res Hum Retrovir*. 1996;12(15):1463-1471.
16. Hayashi T, Hayashi K. Calcium spirulan, an inhibitor of enveloped virus replication, from a Blue-Green Alga *Spirulina platensis*. *J Natural Products*. 1996;59:83-87.
17. Ayehunie S, Belay A, et al. Inhibition of HIV-1 replication by an aqueous extract of *Spirulina platensis*. *J Aquir Immun Defic Syndr Hum Retrovirol*. 1998;18(1):7-12.
18. Schaeffer DJ, Krylov VS. Anti-HIV activity of extracts and compounds from algae and cyanobacteria. *Ecotoxicology and Environmental Safety*. 2000;45:208-227.
19. Mathew B, Sankaranarayanan R, et al. Evaluation of chemo-prevention of oral cancer with *Spirulina fusiformis*. *Nutr Cancer*. 1995;24(2):197-202.
20. Schwartz J. Shklar G. Regression of experimental hamster cancer by beta carotene and algae extracts. *J Oral Maxillofac Surg*. 1987;45:510-515.
21. Schwartz J, Shklar G, et al. Prevention of experimental oral cancer by extracts of *Spirulina-Dunaliella* algae. *Nutr Cancer*. 1988;11:127-134.
22. Mishima T, Murata J, et al. Inhibition of tumor invasion and metastasis by calcium spirulan (Ca-SP), a novel sulfated poly-saccharide derived from a blue-green algae, *Spirulina platensis*. *Clin Exp Metastasis*. 1998;16(6):541-550.
23. Tokuda H, Nishino H, et al. Inhibition of 12-O-tetrade-canoylphorbol-13-acetate promoted mouse skin papilloma by digalactosyl diacylglycerols from the fresh water cyanobacterium *Phormidium tenue*. *Cancer Lett*. 1996;104(1):91-95.
24. Mittal A, Kumar PV, et al. Modulatory potential of *Spirulina fusiformis* on carcinogen metabolizing enzymes in Swiss albino mice. *Phytother Res*. 1999;13(2):111-114.
25. Reddy CM, Bhat VB, et al. Selective inhibition of cyclooxyge-nase-2 by C-phycocyanin, a biliprotein from *Spirulina platensis*. *Biochem Biophys Res Commun*. 2000;277(3):599-603.
26. Bhat VB, Madyastha KM. C-phycocyanin: a potent peroxyl radical scavenger in vivo and in vitro. *Biochem Biophys Res Commun*. 2000;275(1):20-25.
27. Vadiraja BB, Gaikwad NW, Madyastha KM. Hepatoprotective effect of C-phycocyanin: protection for carbon tetrachloride and R-(+)-pulegone-mediated hepatotoxicity in rats. *Biochem Biophys Res Commun*. 1998;249(2):428-431.
28. Romay C, Ledon N, Gonzalez R. Phycocyanin extract reduces leukotriene B4 levels in arachidonic acid-induced mouse-ear inflammation test. *J Pharm Pharmacol*. 1999;51(5):641-642.
29. Gonzalez R, Rodriguez S, et al. Anti-inflammatory activity of phycocyanin extract in acetic acid-induced colitis in rats. *Pharmacol Res*. 1999;39(1):55-59.
30. Kushak RI, Drapeau C, Van Cott EM. Favorable effects of blue-green algae *Aphanizomenon flos-aquae* on rat plasma lipids. *JANA*. 2000;2(3):59-65.
31. Kato T, Takemoto K, et al. Effects of *Spirulina* on dietary hypercholesterolemia in rats. *J Jap Soc Nutr Food Science*. 1984;37:323-332.
32. Hori KG, Ishibashi G, et al. Hypocholesterolemic effect of blue-green alga, ishikurage (*Nostoc commune*) in rats fed atherogenic diet. *Plant Foods Hum Nutr*. 1994;45:63-70.
33. Iwata K, Inayama T, et al. Effects of *Spirulina platensis* on plasma lipoprotein lipase activity in fructose-induced hyper-lipidemic rats. *J Nutr Sci and Vitaminology*. 1999;36:165-171.
34. Nakaya N, Honma Y, et al. Cholesterol lowering effect of *Spirulina* *Nutr Rep Int*. 1988;37:1329-1337.
35. Vlad M, Bordas E, Caseanu E, Uza G, Creteanu E, Polinicenco C. Effect of cuprofilin on experimental athero-sclerosis. *Biol Trace Elem Res*. 1995;48(1):99-109.
36. Becker E, Jakover B, et al. Clinical and biochemical evaluations of the alga *Spirulina* with regard to its application in the treatment of obesity: a double blind cross-over study. *Nutr Rep Int*. 1986;33:565-574.
37. Takai Y, Hosoyamada Y, et al. Effect of water soluble and water insoluble fractions of *Spirulina* over serum lipids and glucose resistance of rats. *J Jap Soc Nutr Food Science*. 1991;44:273-277.
38. Kushak R, VanCott E, Drapeau C, Winter H. Effect of algae *Aphanizomenon flos-aquae* on digestive enzyme activity and polyunsaturated fatty acids level in blood plasma. *Gastroenterol*. 1999;116:A559.
39. Parada JL, Zulpa de Caire G, et al. Lactic acid bacteria growth promoters from *Spirulina platensis*. *Int J Food Microbiol*. 1998;45(3):225-228.
40. Shastri D, Kumar M, Kumar A. Modulation of lead toxicity by *Spirulina fusiformis*. *Phytother Res*. 1999;13(3):258-260.
41. Kapoor R, Mehta U. Supplementary effect of Spirulina on hematological status of rats during pregnancy and lactation. *Plant Foods Hum Nutr*. 1998;52(4):315-324.
42. Valencia A, Walker J. Amulti-axial treatment paradigm for mild traumatic brain injury to achieve reparative functional metaplasticity. 3rd World Congress on Brain Injury, IBIA, Quebec City, June 1999.
43. Drapeau C, Kushak RI, Van Cott EM, Winter HH. Blue-green alga *Aphanizomenon flos-aquae* as a source of dietary polyunsaturated fatty acids and a hypocholesterolemic agent in rats. *J Am Chem Soc*. In press.